

## Distribution of Naturally Occurring Anthraquinones, Iridoids and Flavonoids from *Morinda* Genus: Chemistry and Biological Activity

**Wong PHAKHODEE**

*Department of Chemistry, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand*

(Corresponding author; e-mail: wong\_phakhodee@yahoo.com)

Received: 17 February 2012, Revised: 27 March 2012, Accepted: 9 June 2012

### Abstract

The present review covers chemistry and bioactivities of anthraquinones, iridoids, and flavonoids from the *Morinda* genus. The plants of *Morinda* species, belonging to the Rubiaceae family, have been used as traditional folk medicine with anti-bacterial, anti-fungal, anti-tumor, anti-helmin, analgesic, anti-inflammatory, and immune enhancing effects. They are rich sources of anthraquinones and iridoids. The relevant 2-methoxy-1,3,6-trihydroxyanthraquinone is one of the most potent quinone reductase enzyme inducers with no cytotoxicity with normal cells. Damnacanthol-3-*O*- $\beta$ -D-primeveroside and lucidin-3-*O*- $\beta$ -D-primeveroside displayed a significant reduction of the blood glucose levels in anti-diabetic tests. Additionally, iridoids, 9-*epi*-6 $\alpha$ -methoxy geniposidic acid, scandoside methyl ester, asperulosidic acid, showed a more potent inhibitory effect of melanogenesis than the commercial available depigmented arbutin used in cosmetic industry.

**Keywords:** Rubiaceae, *Morinda*, anthraquinone, iridoid, flavonoid, biological activity

### Introduction

Natural products have continuously served as invaluable resources in terms of searching for structurally novel compounds as leads for the development of drugs having many therapeutic applications. With drug discovery from natural products, it is undeniable that plants are one of the richest sources of natural chemotaxonomy having structural diversity with a broad range of pharmacological activities.

Plants of the genus *Morinda*, classified in the Rubiaceae family, are small evergreen trees or shrubs that consist of about 80 species distributed exclusively in tropical climate zones. An abundance of biologically active and structurally intriguing natural products of *Morinda* species has been widely studied. Almost all parts of these plants including roots, barks, stems, leaves, and fruits, have been used as traditional folk medicine having anti-bacterial, anti-fungal, anti-tumor, anti-helmin, analgesic, anti-inflammatory, and immune enhancing effects [1-6]. The popular product derived from the *Morinda* species, for example,

noni fruit juice (*M. citrifolia*) namely Tahitian Noni® has been approved as a botanical dietary supplement and novel food.

*Morinda* species are well known for the chemical diversities of anthraquinones, iridoids, saccharide fatty acid esters, and lignans. Over 200 compounds have been isolated and identified in the *Morinda* plants. However, chemical composition differs largely depending on the part of the plants. Interestingly, anthraquinones and iridoids with different core frameworks were found as the majority of the genus. The anthraquinones are mainly obtained in the roots of the *Morinda* species, whereas the iridoids are found mainly in the leaves. In addition, flavonoids, coumarins, and triterpenoids were also found in other parts of these plants.

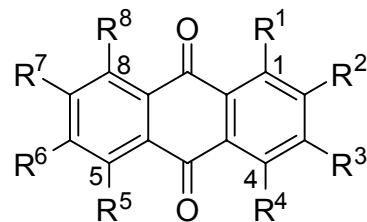
Reviews of *M. citrifolia* including ethnobotany, chemistry, biological activity and safety of the botanical dietary supplement as well as phytochemistry, pharmacology, safety of its fruit have previously been published [7,8].

However, systematic studies of the natural products and their bioactivities from the *Morinda* plants have not yet been reported. The aim of this review is thus to systematically outline the secondary metabolites from *Morinda* species from 1995 to the end of 2011, covering the recent progress on phytochemistry, and biological activity of the naturally occurring anthraquinones, iridoids, and flavonoids.

#### **Anthraquinone from *Morinda* genus**

Anthraquinone is a major component of phytochemical constituents obtained mainly from the roots and fruits of the *Morinda* genus. The distinctive chemical structure of anthraquinones is represented as two aromatic rings connected by two carbonyl carbons. The ring system may be substituted with a variety of alkyl, *O*-alkyl, phenolic, or methoxy groups that give a large variety of possible structures. Either *O*-methoxy or hydroxyl groups of all isolated components from

the *M. species* are mainly located quite uniquely with respect to substitution at C-1 of the anthraquinone core structure as shown in **Table 1**. Recently, the disaccharide anthraquinones such as lucidin-3-*O*- $\beta$ -D-primeveroside **54**, moridone-6-*O*- $\beta$ -D-primeveroside **55**, 1-hydroxy-2-primeverosyloxyethylqnthraquinone-3-olate **60**, 1-hydroxy-5,6-dimethoxy-2-methyl-7-primeverosyloxyanthraquinone **61**, where the disaccharide is located at C-3, C-6, C-2, C-7, respectively, have been reported as depicted in **Figure 1** [9,10].



**Table 1** Anthraquinones isolated from the *Morinda* genus.

No.	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	R <sup>5</sup>	R <sup>6</sup>	R <sup>7</sup>	R <sup>8</sup>
1	H	CH <sub>3</sub>	H	H	H	H	H	H
2	H	COOH	H	H	H	H	H	H
3	H	OCH <sub>3</sub>	H	H	H	H	H	H
4	H	CHO	H	H	H	H	H	H
5	OH	OH	H	H	H	H	H	H
6	OCH <sub>3</sub>	H	OCH <sub>3</sub>	H	H	H	H	H
7	OH	CH <sub>3</sub>	H	H	H	H	H	H
8	OH	CH <sub>2</sub> OH	H	H	H	H	H	H
9	OH	CHO	H	H	H	H	H	H
10	OH	OCH <sub>2</sub> CH <sub>3</sub>	H	H	H	H	H	H
11	CH <sub>3</sub>	H	OH	H	H	H	H	H
12	OCH <sub>3</sub>	OH	H	H	H	H	H	H
13	OCH <sub>3</sub>	CH <sub>3</sub>	H	H	H	H	H	H
14	OCH <sub>3</sub>	H	OH	H	H	H	H	H
15	H	CH <sub>3</sub>	OH	H	H	H	H	H
16	H	CH <sub>2</sub> OH	OH	H	H	H	H	H
17	H	CHO	OH	H	H	H	H	H
18	OH	CH <sub>3</sub>	OH	H	H	H	H	H
19	OCH <sub>3</sub>	CH <sub>3</sub>	OH	H	H	H	H	H
20	OH	CH <sub>2</sub> OH	OH	H	H	H	H	H
21	OH	CH <sub>2</sub> OCH <sub>2</sub> CH <sub>3</sub>	OH	H	H	H	H	H
22	OCH <sub>3</sub>	CH <sub>2</sub> OH	OH	H	H	H	H	H
23	OH	CHO	OH	H	H	H	H	H
24	OH	OH	OH	H	H	H	H	H
25	OH	OH	CH <sub>3</sub>	H	H	H	H	H
26	OCH <sub>3</sub>	CHO	OH	H	H	H	H	H

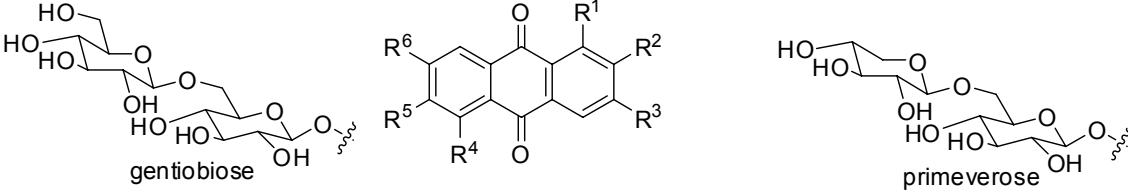
No.	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	R <sup>5</sup>	R <sup>6</sup>	R <sup>7</sup>	R <sup>8</sup>
27	OH	OCH <sub>3</sub>	OH	H	H	H	H	H
28	OH	OCH <sub>3</sub>	OCH <sub>3</sub>	H	H	H	H	H
29	OH	CH <sub>2</sub> O(CH <sub>2</sub> ) <sub>3</sub> CH <sub>3</sub>	OH	H	H	H	H	H
30	OH	CH <sub>2</sub> OCH <sub>3</sub>	OH	H	H	H	H	H
31	OCH <sub>3</sub>	CH <sub>2</sub> OCH <sub>3</sub>	OH	H	H	H	H	H
32	OCH <sub>3</sub>	CH <sub>2</sub> OCH <sub>3</sub>	OCH <sub>3</sub>	H	H	H	H	H
33	OCH <sub>3</sub>	OH	OCH <sub>3</sub>	H	H	H	H	H
34	OCH <sub>3</sub>	OCH <sub>3</sub>	OH	H	H	H	H	H
35	OH	CH <sub>3</sub>	H	H	H	OH	H	H
36	OH	OH	H	H	H	OH	H	H
37	OCH <sub>3</sub>	H	OCH <sub>3</sub>	CH <sub>2</sub> OH	H	H	H	H
38	OH	OCH <sub>3</sub>	OH	H	H	H	H	OH
39	OCH <sub>3</sub>	OH	OCH <sub>3</sub>	H	H	OH	H	H
40	OH	CH <sub>3</sub>	OH	H	H	OH	H	H
41	OH	OCH <sub>3</sub>	OH	H	H	OH	H	H
42	OH	OH	H	H	OH	CH <sub>3</sub>	H	H
43	OH	H	CH <sub>3</sub>	H	H	OCH <sub>3</sub>	H	OH
44	OCH <sub>3</sub>	CH <sub>3</sub>	H	H	OCH <sub>3</sub>	OCH <sub>3</sub>	H	H
45	OCH <sub>3</sub>	OH	H	H	H	H	CH <sub>2</sub> OCH <sub>3</sub>	OCH <sub>3</sub>
46	OH	CH <sub>2</sub> OCH <sub>3</sub>	H	H	OCH <sub>3</sub>	OH	H	H
47	OCH <sub>3</sub>	CH <sub>2</sub> OCH <sub>3</sub>	H	H	OCH <sub>3</sub>	OH	H	H
48	OH	CH <sub>3</sub>	H	H	OCH <sub>3</sub>	OH	H	H
49	OH	CH <sub>3</sub>	H	H	OH	OCH <sub>3</sub>	H	H
50	OH	CH <sub>2</sub> OH	H	H	OCH <sub>3</sub>	H	H	OH
51	OH	CH <sub>2</sub> OCH <sub>3</sub>	H	H	OH	OCH <sub>3</sub>	OH	H
52	OH	CH <sub>3</sub>	OCH <sub>3</sub>	H	H	H	OCH <sub>3</sub>	OH

### Biological activity of anthraquinone from the *Morinda* genus

The genus *Morinda* is loaded with different types of anthraquinones and its congener. The potential pharmacological activities of the anthraquinones and its derivative have been published. These pertain to antibacterial, anti-cancer, anti-inflammatory, antioxidant, cancer-chemoprevention, anti-microbial activity, and anti-osteoporotic activity. Anthraquinones from the *Morinda* plants and their bioactivities are summarized in **Table 2**.

The effects of anthraquinones on the growth of H1299 human lung cancer cells, HCT116 human colon adenocarcinoma cells, KB human mouth epidermal carcinoma cells, and HeLa human cervical carcinoma cells have been investigated [2,6]. Anthraquinones **4** and **7** showed potent growth-inhibitory effect on HCT116 with IC<sub>50</sub> values of 5.9 µg/mL and 6.9 µg/mL, respectively. H1299 has been inhibited by anthraquinones **4**, **7**, and **12** with IC<sub>50</sub> values of

4.9, 4.1, 4.3 µg/mL, respectively. In KB cell lines, anthraquinones **23**, **26**, and **35** have been reported to display moderate growth inhibitory activity with IC<sub>50</sub> values of 5.99, 6.35, 7.68 µg/mL and showed weak cytotoxicity against HeLa human cervical carcinoma cells. Additionally, anthraquinones **18**, **23**, **26**, and **30** possessed strong activity towards the CEM-SS cell line with CC<sub>50</sub> of 3, 1.7, 4 and 3 µg/mL, respectively. The anthraquinones **26** and **30** also showed strong cytotoxicity towards MCF-7 cell line with the same CC<sub>50</sub> value of 3 µg/mL. The significant cytotoxicity against CEM-SS and MCF-7 cell lines was probably due to the combination of the differently substituted types at C-1 to C-3, especially the hydroxylated at C-1 and C-3. Moreover, the presence of the formyl group at C-2 and hydroxylated moiety at C-3 of nordamnacanthal **23** and damnacanthal **26** was found to enhance the anthraquinones microbial activity [11].



	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	R <sup>5</sup>	R <sup>6</sup>
<b>53</b>	OCH <sub>3</sub>	CH <sub>2</sub> OH	CH <sub>2</sub> O-primeverose	H	H	H
<b>54</b>	OH	CH <sub>2</sub> OH	CH <sub>2</sub> O-primeverose	H	H	H
<b>55</b>	OH	CH <sub>3</sub>	H	OH	O-primeverose	H
<b>56</b>	OCH <sub>3</sub>	CH <sub>2</sub> O-gentiobiose	H	H	H	H
<b>57</b>	OH	CH <sub>2</sub> O-primeverose	H	H	H	H
<b>58</b>	OCH <sub>3</sub>	CH <sub>2</sub> O-primeverose	OH	H	H	H
<b>59</b>	OCH <sub>3</sub>	CH <sub>2</sub> O-primeverose	O <sup>-</sup>	H	H	H
<b>60</b>	OH	CH <sub>2</sub> O-primeverose	O <sup>-</sup>	H	H	H
<b>61</b>	OH	CH <sub>3</sub>	H	OCH <sub>3</sub>	OCH <sub>3</sub>	O-primeverose

**Figure 1** Glycoside anthraquinones isolated from *Morinda* species.

A quinone reductase (QR) bioassay is one of the strategies of cancer chemoprevention for protecting cells from carcinogenesis. This is involved in the catalytic oxidation of NADH or NADPH and consequently deactivates the amount of harmful radicals and electrophiles in phase II metabolizing enzyme [3,5]. With the evaluation of anthraquinones **5**, **12**, **16**, **18**, **27**, **40**, **41**, **48**, and **45**, compounds **5**, **16**, **18**, **40**, **41**, and **50** exhibited QR induction activity with CD values of 12, 0.94, 8.1, 0.56, 0.009, and 1.67 µM, respectively. 2-Methoxy-1,3,6-trihydroxyanthraquinone **41** was found to be potent inducer of QR activity (0.009 µM) which was higher than L-sulforaphane (0.34 µM), a standard inducer, a well-known cancer chemopreventive agent isolated from broccoli [12]. No evidence of cytotoxicity was observed at up to

20 µg/mL (> 69.9 µM). It should be noted that the difference in substitutions pattern at C-1, C-2, C-3 and C-6, especially at C-2 position play an important role in both potency and cytotoxicity in the QR induction assay [3,5].

The tests of three anthraquinones, **53-55**, and two iridoids, **63** and **67**, for hypoglycemic effect have also been investigated. Anthraquinones **53** and **54** displayed a significant reduction of the blood glucose levels at 5 h after administration, but anthraquinone **55** and two iridoids, **63** and **67** have no effects with reducing blood glucose level. Particularly, the anthraquinone having no substituents on one aromatic ring seems to have a significant influence on the hypoglycemic effect [9].

**Table 2** Biological activities of naturally occurring anthraquinones from the *Morinda* genus.

Structural name of anthraquinone	<i>Morinda</i> specie(s)	Plant part	Biological activity	Ref
Tectoquinone <b>1</b>	<i>M. lucida</i>	Roots	-	[13]
Anthraquinone-2-carboxylic acid <b>2</b>	<i>M. officinalis</i>	Roots	-	[14]
2-Methoxyanthraquinone <b>3</b>	<i>M. officinalis</i>	Roots	-	[15]
2-Formylanthraquinone <b>4</b>	<i>M. citrifolia</i>	Roots	H1299 <sup>a</sup> ( $IC^{e}_{50} = 4.9 \pm 1.2 \mu\text{g/mL}$ ) HT116 <sup>b</sup> ( $IC_{50} = 5.9 \pm 1.5 \mu\text{g/mL}$ )	[2]
1,2-Dihydroxyanthraquinone <b>5</b>	<i>M. citrifolia</i>	Fruits	-	[16]
	<i>M. citrifolia</i>	Roots	QR <sup>c</sup> induction assay ( $CD^d = 12.0 \mu\text{M}$ , $IC_{50} = 14.9 \mu\text{g/mL}$ )	[5]
1,3-Dimethoxyanthraquinone <b>6</b>	<i>M. citrifolia</i>	Fruits	-	[16]
1-Hydroxy-2-methylanthraquinone <b>7</b>	<i>M. lucida</i>	Roots	-	[13,14]
	<i>M. citrifolia</i>	Roots	H1299 ( $IC_{50} = 4.1 \pm 1.2 \mu\text{g/mL}$ ) HT116 ( $IC_{50} = 6.9 \pm 1.7 \mu\text{g/mL}$ )	[2]
Digiferruginol <b>8</b>	<i>M. citrifolia</i>	Roots	Potential larvicultural of <i>Aedes aegypti</i>	[17]
2-Formyl-1-hydroxyanthraquinone <b>9</b>	<i>M. officinalis</i>	Roots	-	[14]
	<i>M. citrifolia</i>	Roots	-	[2,17]
	<i>M. elliptica</i>	Roots	-	[18]
2-Ethoxy-1-hydroxyanthraquinone <b>10</b>	<i>M. citrifolia</i>	Roots	-	[17]
1-Methyl-3-hydroxyanthraquinone <b>11</b>	<i>M. citrifolia</i>	Roots	-	[2]
Alizarin-1-methyl ether <b>12</b>	<i>M. lucida</i>	Roots	<i>C. cucumerinum</i> <sup>f</sup> (0.5 $\mu\text{g}$ ) <i>C. albicans</i> <sup>f</sup> (1.0 $\mu\text{g}$ )	[13]
	<i>M. officinalis</i>	Roots	-	[14,15]
	<i>M. citrifolia</i>	Roots	H1299 ( $IC_{50}=4.3 \pm 1.5 \mu\text{g/mL}$ )	[2]
	<i>M. citrifolia</i>	Fruits	-	[3,19]
	<i>M. elliptica</i>	<sup>g</sup> Cell Cul	Antioxidant [FCT method] (comparable to $\alpha$ -tocopherol)	[4]
1-Methoxy-2-methylanthraquinone <b>13</b>	<i>M. pandurifolia</i>	Roots	-	[6]
1-Methoxy-3-hydroxyanthraquinone <b>14</b>	<i>M. citrifolia</i>	Roots	-	[2]
3-Hydroxy-2-methylanthraquinone <b>15</b>	<i>M. officinalis</i>	Roots	-	[14]
3-Hydroxy-2-hydroxymethyl-anthraquinone <b>16</b>	<i>M. lucida</i>	Roots	-	[13]
	<i>M. officinalis</i>	Roots	-	[15]
	<i>M. citrifolia</i>	Roots	QR induction assay ( $CD = 0.94 \mu\text{M}$ , $IC_{50}>20 \mu\text{g/mL}$ )	[5]
3-Hydroxyanthraquinone-2-carbaldehyde <b>17</b>	<i>M. lucida</i>	Roots	<i>C. cucumerinume</i> (2.0 $\mu\text{g}$ ) <i>C. albicanse</i> (5.0 $\mu\text{g}$ )	[13]
Rubiadin <b>18</b>	<i>M. officinalis</i>	Roots	-	[14]
	<i>M. citrifolia</i>	Roots	-	[2]
	<i>M. elliptica</i>	Cell Cul	-	[4]
	<i>M. citrifolia</i>	Roots	QR induction assay	[5]
	<i>M. angustifolia</i>	Roots	( $CD = 8.1 \mu\text{M}$ , $IC_{50}>20 \mu\text{g/mL}$ )	[20]
	<i>M. elliptica</i>	Roots	-	[20]
Rubiadin-1-methyl ether <b>19</b>	<i>M. lucida</i>	Roots	<sup>h</sup> CEM-SS cells ( $CC^{i}_{50} = 1.7 \mu\text{g/mL}$ )	[11]
Lucidin <b>20</b>	<i>M. officinalis</i>	Roots	-	[13]
	<i>M. officinalis</i>	Roots	-	[14,15]
Ibericin <b>21</b>	<i>M. pandurifolia</i>	Roots	-	[14]
	<i>M. officinalis</i>	Roots	-	[6]
	<i>M. citrifolia</i>	Roots	-	[14]
	<i>M. angustifolia</i>	Roots	-	[2]
Damnacanthol <b>22</b>	<i>M. lucida</i>	Roots	-	[20]
	<i>M. angustifolia</i>	Roots	-	[13]
	<i>M. angustifolia</i>	Roots	-	[20]

Structural name of anthraquinone	<i>Morinda</i> specie(s)	Plant part	Biological activity	Ref
Nordamnacanthal <b>23</b>	<i>M. lucida</i>	Roots	<i>C. cucumerinum</i> (1.0 µg) <i>C. albicans</i> (1.0 µg)	[13]
	<i>M. citrifolia</i>	Roots	-	[2,17]
	<i>M. elliptica</i>	Cell Cul	Antioxidant [FCT method] (stronger than $\alpha$ -tocopherol)	[4]
	<i>M. pandurifolia</i>	Roots	EBV <sup>j</sup> activation in Raji cells (IC <sub>50</sub> = 0.4 µg/mL) inhibition rate	
	<i>M. elliptica</i>	Roots	75.0% cell viability 75.8%	
	<i>M. pandurifolia</i>	Roots	KB <sup>k</sup> cells (IC <sub>50</sub> = 5.99 µg/mL)	[6]
	<i>M. elliptica</i>	Roots	CEM-SS cells (CC <sub>50</sub> = 1.7 µg/mL)	[11]
Anthragallol <b>24</b>	<i>M. pandurifolia</i>	Roots	-	[6]
1,2-Dihydroxy-3-methylanthraquinone <b>25</b>	<i>M. officinalis</i>	Roots	-	[15]
Damnacanthal <b>26</b>	<i>M. lucida</i>	Roots	-	[13]
	<i>M. citrifolia</i>	Roots	-	[2]
	<i>M. citrifolia</i>	Roots	Potential larvicultural of <i>Aedes aegypti</i>	[17]
	<i>M. pandurifolia</i>	Roots	KB cells (IC <sub>50</sub> =6.35 µg/mL)	
	<i>M. elliptica</i>	Roots	CEM-SS cells (CC <sub>50</sub> = 4 µg/mL)	[6]
	<i>M. elliptica</i>	Roots	MCF-7 <sup>l</sup> cells (CC <sub>50</sub> = 3 µg/mL)	[11]
Anthragallol-2-methyl ether <b>27</b>	<i>M. officinalis</i>	Roots	-	[14]
	<i>M. citrifolia</i>	Fruits	-	[3,19]
	<i>M. citrifolia</i>	Fruits	EBV-EA <sup>m</sup> induction (IC <sub>50</sub> =483 mol ratio/32 pmol TPA)	[21]
Anthragallol-2,3-dimethyl ether <b>28</b>	<i>M. pandurifolia</i>	Roots	-	[6]
Lucidin- $\omega$ -butyl ether <b>29</b>	<i>M. angustifolia</i>	Roots	-	[20]
Lucidin- $\omega$ -methyl ether <b>30</b>	<i>M. elliptica</i>	Cell Cul	Antioxidant [FCT method] (comparable to $\alpha$ -tocopherol)	[4]
	<i>M. pandurifolia</i>	Roots	-	[6]
	<i>M. elliptica</i>	Roots	CEM-SS cells (CC <sub>50</sub> = 3 µg/mL)	[11]
	<i>M. elliptica</i>	Roots	MCF-7 cells (CC <sub>50</sub> = 3 µg/mL)	[11]
3-Hydroxy-1-methoxy-2-methoxymethylanthraquinone <b>31</b>	<i>M. pandurifolia</i>	Roots	-	[6]
1,3-Dimethoxy-2-methoxymethylanthraquinone <b>32</b>	<i>M. citrifolia</i>	Roots	-	[2]
Anthragallol-1,3-dimethyl ether <b>33</b>	<i>M. citrifolia</i>	Fruits	-	[19]
1,2-Dimethoxy-3-hydroxyanthraquinone <b>34</b>	<i>M. officinalis</i>	Roots	-	[14]
Soranjidiol <b>35</b>	<i>M. elliptica</i>	Cell Cul	-	[4]
	<i>M. pandurifolia</i>	Roots	KB cells (IC <sub>50</sub> =7.67 µg/mL)	[6]
Flavopurpurin <b>36</b>	<i>M. pandurifolia</i>	Roots	-	[6]
Morindicininone <b>37</b>	<i>M. citrifolia</i>	Stems	-	[22]
1,3,8-Trihydroxy-2-methoxyanthraquinone <b>38</b>	<i>M. officinalis</i>	Roots	Strong inhibitory effect on osteoclastic bone resorption	[15]
6-Hydroxy-anthragallol-1,3-dimethylether <b>39</b>	<i>M. citrifolia</i>	Fruits	-	[19]
1,3,6-Trihydroxy-2-methylantraquinone <b>40</b>	<i>M. citrifolia</i>	Roots	QR induction assay (CD=0.56 µM, IC <sub>50</sub> =12.8 µg/mL)	[5]
2-Methoxy-1,3,6-trihydroxyanthraquinone <b>41</b>	<i>M. citrifolia</i>	Fruits	QR induction assay (CD=0.009 µM, IC <sub>50</sub> >20 µg/mL)	[3]
Morindone <b>42</b>	<i>M. lucida</i>	Roots	-	[13]
	<i>M. elliptica</i>	Cell cul	Antioxidant[DPPH assay] ( IC <sub>50</sub> =40.6 µg/mL)	[4]
			Antioxidant [FCT method] (stronger than $\alpha$ -tocopherol)	
Physcion <b>43</b>	<i>M. officinalis</i>	Roots	-	[14]

Structural name of anthraquinone	<i>Morinda</i> specie(s)	Plant part	Biological activity	Ref
1,5,6-Trimethoxy-2-methylanthraquinone <b>44</b>	<i>M. lucida</i>	Roots	Strong inhibitory effect on osteoclastic bone resorption	[15]
Morindicinone <b>45</b>	<i>M. citrifolia</i>	Stems	-	[13]
5,15-di- <i>O</i> -methylmorindol <b>46</b>	<i>M. citrifolia</i>	Fruits	-	[22]
			EBV-EA induction (IC <sub>50</sub> =475 mol ratio/32 pmol TPA)	[19]
			-	[21]
1,5,15-tri- <i>O</i> -methylmorindol <b>47</b>	<i>M. citrifolia</i>	Fruits	EBV-EA induction (IC <sub>50</sub> = 386 mol ratio/32 pmol TPA)	[23]
Morindone-5-methylether <b>48</b>	<i>M. citrifolia</i>	Leaves	-	[21]
Morindone-6-methylether <b>49</b>	<i>M. citrifolia</i>	Fruits	-	[24]
1,8-Dihydroxy-2-hydroxymethyl-5-methoxyanthraquinone <b>50</b>	<i>M. citrifolia</i>	Roots	-	[3,18]
1,5,7-Trihydroxy-6-methoxy-2-methoxymethylanthraquinone <b>51</b>	<i>M. citrifolia</i>	Fruits	QR induction assay (CD=1.67 μM, IC <sub>50</sub> >20 μg/mL)	[17]
1,8-Dihydroxy-2-methyl-3,7-dimethoxyanthraquinone <b>52</b>	<i>M. angustifolia</i>	Fruits	-	[3]
Damnacanthol-3- <i>O</i> -β-D-primeveroside <b>53</b>	<i>M. citrifolia</i>	Roots	Antimicrobial activities against Zone of inhibition (diameter, mm) <i>B. subtilis</i> (14.0 mm, with 13.3μg/disc) <i>E. coli</i> (12.5 mm, with 13.3μg/disc) <i>M. luteus</i> (13.0 mm, with 13.3μg/disc) <i>S. lutea</i> (6.8 mm, with 13.3μg/disc) <i>C. albicans</i> (7.5 mm, with 13.3μg/disc) <i>S. sake</i> (6.3 mm, with 13.3μg/disc)	[20]
Lucidin-3- <i>O</i> -β-D-primeveroside <b>54</b>	<i>M. angustifolia</i>	Roots	Reduction of blood glucose level	[9]
	<i>M. citrifolia</i>	Roots	from streptozotocan-induced diabetic mice	
Morindone-6- <i>O</i> -β-D-primeveroside <b>55</b>	<i>M. citrifolia</i>	Roots	-	[9]
Digiferruginol-1-methylether-11- <i>O</i> -β-gentibioside <b>56</b>	<i>M. citrifolia</i>	Roots	-	[10]
Degiferruginol-11-O-β -primeverside <b>57</b>	<i>M. citrifolia</i>	Roots	-	[10]
Damnacanthol-11-O-β -primeverside <b>58</b>	<i>M. citrifolia</i>	Roots	-	[10]
1-Methoxy-2-primeverosyloxymethylanthraquinone-3-olate <b>59</b>	<i>M. citrifolia</i>	Roots	-	[10]
1-Hydroxy-2-primeverosyloxymethylanthraquinone-3-olate <b>60</b>	<i>M. citrifolia</i>	Roots	-	[10]
1-Hydroxy-5,6-dimethoxy-2-methyl-7-primeverosyloxyanthraquinone <b>61</b>	<i>M. citrifolia</i>	Roots	-	[10]

<sup>a</sup>Human lung cancer lines (H1299) <sup>b</sup>Human colon adenocarcinoma cell lines (HT116) <sup>c</sup>Quinone Reductase (QR) induction assay <sup>d</sup>Concentration required to double QR activity (CD) <sup>e</sup>Concentration inhibiting cell growth by 50 % (IC<sub>50</sub>) <sup>f</sup>Bioautography assay <sup>g</sup>Cell cult = Cell Culture <sup>h</sup>T-lymphoblastic leukaemia (CEM-SS cells) <sup>i</sup>50 % cytotoxic concentration (CC<sub>50</sub>) <sup>j</sup>Epstien Barr Virus activation on Raji cells (EBV activation on Raji cells) <sup>k</sup>Human mouth epiderma carcinoma cell lines (KB) <sup>l</sup>Breast carcinoma (MCF-7 cells) <sup>m</sup>TPA-induced inflammation in mice and on the induction of Epstein-Barr Virus Early Antigen.

### Chemistry of iridoid from *Morinda* genus

Iridoid is a monoterpenoid containing cyclopentane ring of iridane skeleton usually fused with a pyran ring. *Morinda* plant leaves were found to be the richest sources of diverse monoterpene iridoids as shown in **Figures 2** and **3**. Based on structural analysis, the majority of these iridoids contains monosaccharides connected to C-1 and either methyl ester or carboxylic acid at C-4. The fused ring junction was observed as *cis*-geometry except for 9-*epi*-6 $\alpha$ -methoxy geniposidic acid **64** where there is a *trans*-geometry [25]. The complicated structure based on the stereogenic center of the iridoids **83 - 96** was analyzed mainly as spirolactone molecules as depicted in **Figure 3**, and several of them have been revised by Schripsema *et al.* [26]. In addition, the structural revision of the 6-5 fused ring of moridacin **62** to 5-5 fused ring of borreriagenin **97** was also reviewed [26].

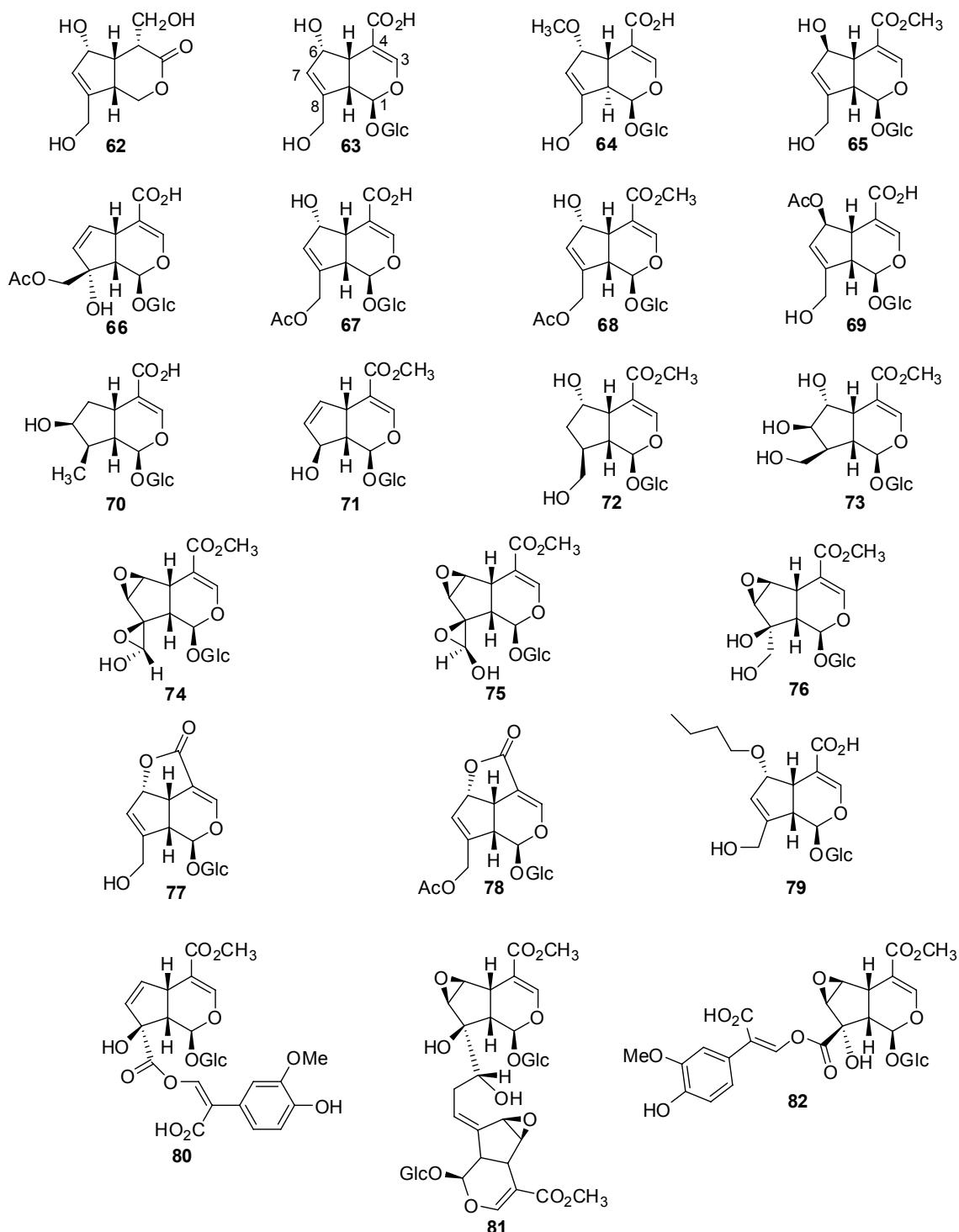
### Biological activity of iridoid from *Morinda* genus

The promising potential pharmacological studies of iridoids revealed anti-melanogenesis activity, UVB-induced AP-1 activity, anti-complementary activity, and anti-malarial activity against *Plamodium falciparum*. Iridoids from the *Morinda* plants and their bioactivities are summarized in **Table 3**.

Transcriptional activator protein-1 (AP-1) plays as key role in the development of human skin-cancer by irradiation of UVB. The inhibition of the AP-1 activity has been shown to suppress cell transformation and tumor promotion. Citrifolinin A **80** and citrifolinoside **82** showed significant inhibitory effect with IC<sub>50</sub> values of 69.6 and 29.0  $\mu$ M, respectively [35,36].

Interestingly, although iridoids **84 - 87** and **89** had weak cytotoxicity against KB cell line, they showed more potential inhibitory for the proliferation of the malarial parasite (*P. falciparum*). In particular, dehydromethoxy-gaertneroside **89** showed superior inhibition with an IC<sub>50</sub> 0.04  $\mu$ M without cytotoxicity [38]. The functionalities of 6"-acetyl, 3'-methoxy and 7'-ketonic carbonyl groups seem to play an important role in enhancing anti-malarial activity independent of cytotoxicity against the host cells. The iridoids **84** and **85** were also observed for anti-complementary activity with IC<sub>50</sub> values of 58 and 71  $\mu$ M, respectively.

Iridoids **64**, **65**, **67** showed remarkable inhibition of melanogenesis. These compounds were found to exhibit more potent inhibitory effect of melanogenesis than those of arbutin which is recognized as a useful depigmentation substrate for the skin whitening in the cosmetic industry [25,41].



**Figure 2** Phytochemical constituents of iridoids isolated from the *Morinda* species.

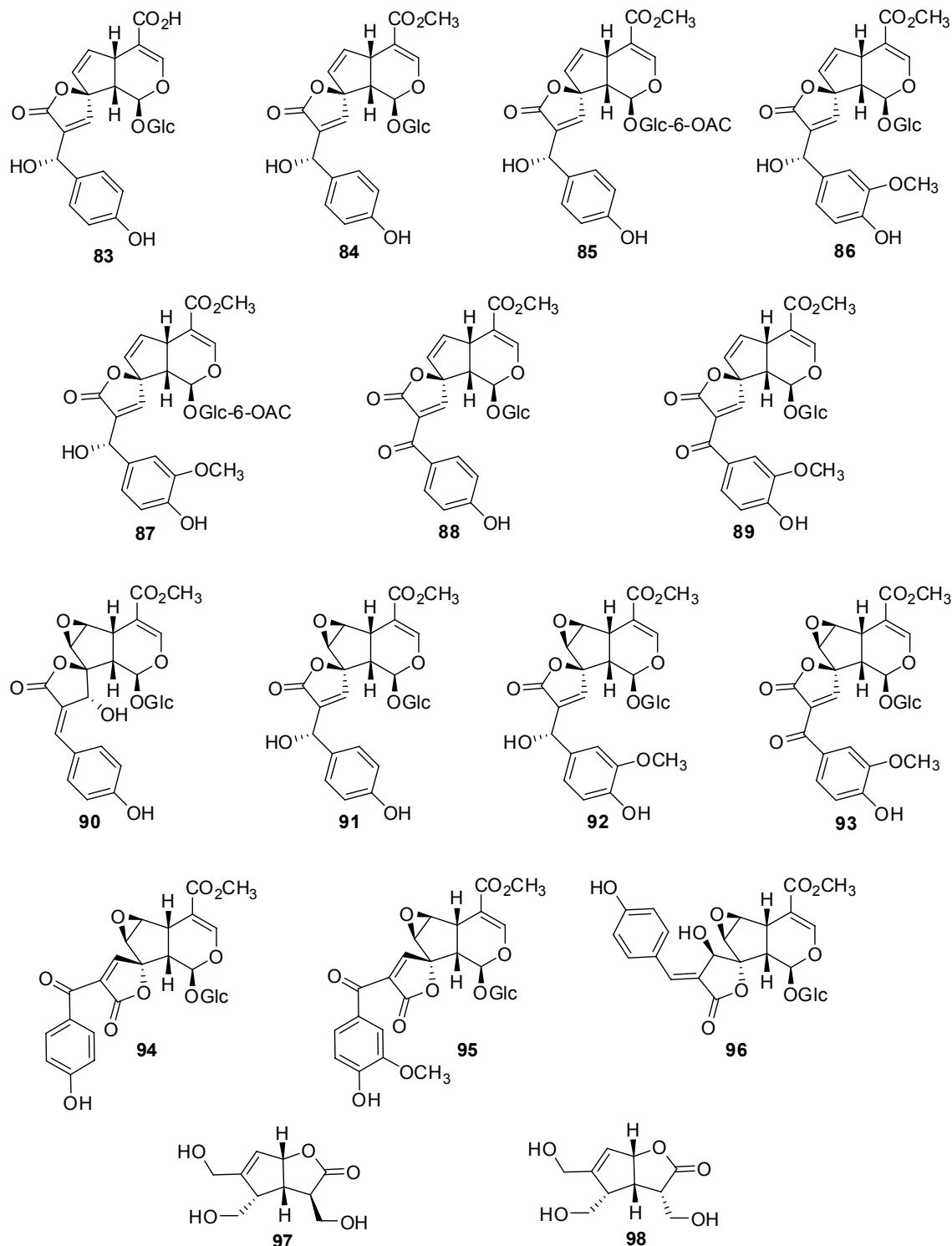


Figure 3 Spirolactone molecules and 5-5 fused ring iridoids isolated from the *Morinda* species.

**Table 3** Biological activities of naturally occurring iridoids from *Morinda* genus.

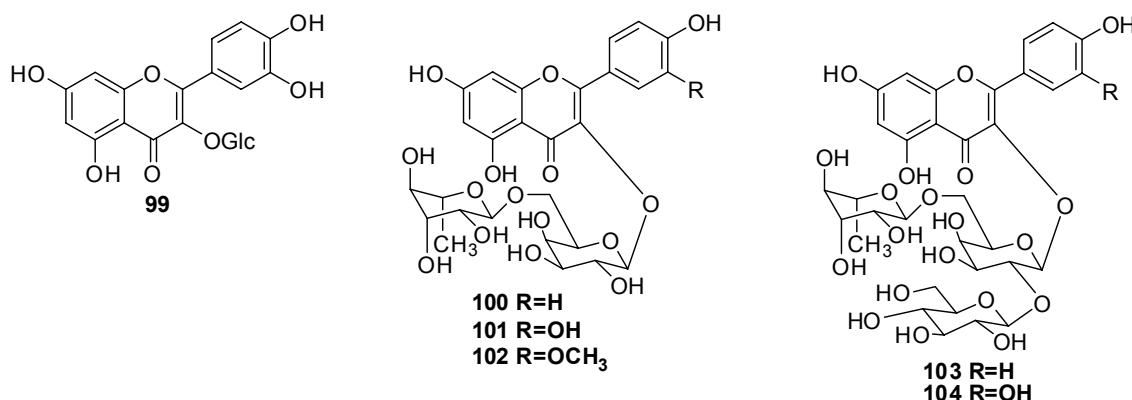
Structural name of iridoid	<i>Morinda</i> specie(s)	Plant part	Biological activity	Ref
Morindacin <b>62</b>	<i>M. citrifolia</i>	Fruits	-	[19]
Deacetylasperulosidic acid <b>63</b>	<i>M. citrifolia</i>	Fruits	-	[19,25]
	<i>M. citrifolia</i>	Fruits	-	[27]
		juice		
9- <i>epi</i> -6 $\alpha$ -methoxy geniposidic acid <b>64</b>	<i>M. citrifolia</i>	Fruits	Antimelanogenesis activity (38 % reduce melanin content at 100 $\mu$ M)	[25]
Scandoside methyl ester <b>65</b>	<i>M. citrifolia</i>	Fruits	Antimelanogenesis activity (46 % reduce melanin content at 100 $\mu$ M)	[25]
10- <i>O</i> -Acetylmonotropein <b>66</b>	<i>M. coreia</i>	Leaves and branches	-	[28]
Asperulosidic acid <b>67</b>	<i>M. citrifolia</i>	Fruits	EBV-EA induction ( $IC_{50}$ = 3485 mol ratio/32 pmol TPA)	[21]
	<i>M. citrifolia</i>	Fruits	Inhibition of tumorigenesis	[29]
	<i>M. citrifolia</i>	Fruits	Antimelanogenesis activity (45 % reduce melanin content at 100 $\mu$ M)	[25]
	<i>M. elliptica</i>	Leaves, branches	-	[30]
Asperulosidic acid methyl ester <b>68</b>	<i>M. citrifolia</i>	Fruits, leaves	-	[31]
6- <i>O</i> -acetylcandoside <b>69</b>	<i>M. coreia</i>	Leaves, branches	-	[28]
Loganic acid <b>70</b>	<i>M. citrifolia</i>	Seeds	-	[32]
Citrifoside <b>71</b>	<i>M. citrifolia</i>	Leaves	-	[24]
6 $\alpha$ -Hydroxyadoxoside <b>72</b>	<i>M. citrifolia</i>	Fruits	-	[33]
Yopaaoside C <b>73</b>	<i>M. coreia</i>	Leaves, branches	-	[28]
Citrifolinin Ba <b>74</b>	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] ( $IC_{50}$ =30 $\mu$ M, 7.7 %)	[34]
Citrifolinin Bb <b>75</b>	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] ( $IC_{50}$ =30 $\mu$ M, 7.7 %)	[34]
6 $\beta$ ,7 $\beta$ -epoxy-8- <i>epi</i> -splendoside <b>76</b>	<i>M. citrifolia</i>	Fruits	-	[14]
Deacetylasperuloside <b>77</b>	<i>M. citrifolia</i>	Fruits, leaves	-	[31]
Asperuloside <b>78</b>	<i>M. elliptica</i>	Leaves, branches	-	[30]
Rhodolatouside <b>79</b>	<i>M. citrifolia</i>	Seeds	-	[32]
Citrifolinin A <b>80</b>	<i>M. citrifolia</i>	Leaves	UVB-induced AP-1 activity ( $IC_{50}$ =69.6 $\mu$ M)	[35,36]
Citrifolinin A-1 <b>81</b>	<i>M. citrifolia</i>	Leaves	UVB-induced AP-1 activity ( $IC_{50}$ =29.0 $\mu$ M)	[36,40]
Citrifolinoside <b>82</b>	<i>M. citrifolia</i>	Leaves	-	
Gaertneric acid <b>83</b>	<i>M. morindoides</i>	Leaves	Anticomplementary activity ( $IC_{50}$ =69 $\mu$ M)	[37]
Gaertneroside <b>84</b>	<i>M. morindoides</i>	Leaves	Anticomplementary activity ( $IC_{50}$ =58 $\mu$ M), Anti-malarial activity ( <i>P. falciparum</i> ) ( $IC_{50}$ =0.8 $\mu$ M)	[37]
				[38]

Structural name of iridoid	<i>Morinda</i> specie(s)	Plant part	Biological activity	Ref
6-Acetylgaertneroside <b>85</b>	<i>M. morindoides</i>	Leaves	Anticomplementary activity (IC <sub>50</sub> =71 μM) Anti-malarial activity ( <i>P.falciparum</i> ) (IC <sub>50</sub> =4.1 μM)	[37]
				[38]
Methoxygaertneroside <b>86</b>	<i>M. morindoides</i>	Leaves	Anti-malarial activity ( <i>P.falciparum</i> ) (IC <sub>50</sub> =21.9 μM)	[38]
6-Acetylmethoxygaertneroside <b>87</b>	<i>M. morindoides</i>	Leaves	Anti-malarial activity ( <i>P.falciparum</i> ) (IC <sub>50</sub> =0.1 μM)	[38]
Dehydrogaertneroside <b>88</b>	<i>M. morindoides</i>	Leaves	-	[37]
Dehydromethoxygaertneroside <b>89</b>	<i>M. morindoides</i>	Leaves	-	[37]
			Anti-malarial activity ( <i>P.falciparum</i> ) (IC <sub>50</sub> =0.04 μM)	[38]
Citrifolinoside A <b>90</b>	<i>M. citrifolia</i>	Leaves	Revision of structure <b>80</b>	[26]
Epoxygaertneroside <b>91</b>	<i>M. morindoides</i>	Leaves	Revision of structure <b>96</b>	[26,39]
Epoxymethoxygaertneroside <b>92</b>	<i>M. morindoides</i>	Leaves	-	[37]
Dehydroepoxymethoxy-gaertneroside <b>93</b>	-	-	-	[28]
Morinipticoside <b>94</b>	<i>M. elliptica</i>	Leaves and branches	Revision of structures <b>82, 95</b>	[26]
			-	[30]
Yopaaoside A <b>95</b>	<i>M. coreia</i>	Leaves and branches	-	[28,30]
Yopaaoside B <b>96</b>	<i>M. elliptica</i> <i>M. coreia</i>	Leaves and branches	-	[28,30]
Borreriagenin <b>97</b>	-	-	Revision of structure <b>62</b>	[26]
4-epi-Borreriagenin <b>98</b>	<i>M. citrifolia</i>	Fruit juice	-	[27]

### Chemistry and biological activity of flavonoids from the *Morinda* genus

Flavonoids, derived from 2-phenylchromen-4-one, represent a highly diverse class of polyphenolic secondary metabolites which are usually abundant in natural products of higher plant origin, but have been reported as rare case of flavonol glycoside from the *Morinda* species. According to **Figure 4** and **Table 4**, the polar flavonol glycosides **99 - 104** containing flavonoid framework, have been isolated from leaves of *M. citrifolia* and are widely known as natural anti-oxidants in vitro [21,33-34]. Compounds **99 - 101, 103**, and **104** showed the antioxidant ability to scavenge DPPH at the concentration of 30 μM with 85.8, 4.5, 79.9, 28.6, and 81.3 %, respectively. Compound **102** was found to be

inactive in DPPH assay, but it was found to be a potent anti-oxidant activity against both authentic ONOO<sup>-</sup> and SIN-1-derivatived ONOO<sup>-</sup>. Narcissoside **102** having a methoxy group at C-3' was also found to be inactive in DPPH assay whereas quercetin derivatives **99, 101, 104** having a hydroxyl group at C-3' displayed a strong anti-oxidant activity [33,34]. Quercetin derivative **101** also exhibited the inhibitory effect on EBA-EA activation induced by TPA compared with those of the known quercetin inhibitor as well as β-carotene. It was found that compound **101** showed comparable activity (578 mol ratio/32pmol TPA) with known quercetin (560 mol ratio/32pmol TPA), but less inhibitory effect than β-carotene (397 mol ratio/32pmol TPA) [21].



**Figure 4** Flavonol glycosides isolated from the *Morinda* species.

**Table 4** Biological activities of naturally occurring flavonol glycosides from *Morinda* genus.

Structural name of flavonol glycoside	<i>Morinda</i> specie(s)	Plant part	Biological activity	Ref
Quercetin-3-O-β-D-glucopyranoside <b>99</b>	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] (IC <sub>50</sub> =30 μM with 85.8 %)	[34]
Kaempferol-3-O-α-L-rhamnopyranosyl-(1→6)-β-D-glucopyranoside <b>100</b>	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] (IC <sub>50</sub> =30 μM with 4.5 %)	[34]
Quercetin-3-O-α-L-rhamnopyranosyl-(1→6)-β-D-glucopyranoside <b>101</b>	<i>M. citrifolia</i>	Fruits	EBV-EA induction (IC <sub>50</sub> =578 mol ratio/32 pmol TPA)	[21]
	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] ( IC <sub>50</sub> =30 μM with 79.9 %)	[34]
Narcissoside <b>102</b>	<i>M. citrifolia</i>	Fruits	Antioxidant [peroxynitrite assay] Authentic ONOO <sup>-</sup> (IC <sub>50</sub> =3.8 μM) SIN-1-derivatived ONOO <sup>-</sup> (IC <sub>50</sub> =9.6 μM)	[33]
Kaempferol-3-O-β-D-glucopyranosyl-(1→2)-[α-L-rhamnopyranosyl-(1→6)]-β-D-galacopyranoside <b>103</b>	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] (IC <sub>50</sub> =30 μM with 28.6 %)	[34]
Quercetin-3-O-β-D-glucopyranosyl-(1→2)-[α-L-rhamnopyranosyl-(1→6)]-β-D-galacopyranoside <b>104</b>	<i>M. citrifolia</i>	Leaves	Antioxidant[DPPH assay] (IC <sub>50</sub> =30 μM with 81.3 %)	[34]

## Conclusion

The phytochemistry of the *Morinda* species has been extensively investigated revealing that they contain anthraquinones, iridoids, flavonoids and other secondary metabolites. These compounds especially anthraquinones, iridoids, flavonoids have been shown to exhibit a wide range of biological activities including antioxidant, anti-malarial, anti-tumor, anti-melanogenesis, anti-diabetic, and chemopreventive activities. In addition, the results from quinone reductase (QR) bioassay gave a promising warrant for further research, especially 2-methoxy-1,3,6-trihydroxyanthraquinone **41** which exhibits a higher potent inducer of QR activity (0.009 µM) than the standard L-sulforaphane.

## Acknowledgements

I would like to thank Asst. Professor Dr. Surat Laphookhieo for his expertise in revising this manuscript.

## References

- [1] W Mian-Ying, BJ West, CJ Jensen, D Nowicki, S Chen, A Palu and G Anderson. *Morinda citrifolia* (noni): A literature review and recent advances in noni research. *Acta Pharmacol. Sin.* 2002; **12**, 1127-41.
- [2] L Lv, H Chen, C-T Ho and S Sang. Chemical components of the roots of noni (*Morinda citrifolia*) and their cytotoxic effects. *Fitoterapia* 2011; **82**, 704-8.
- [3] AD Pawlus, BN Su, WJ Keller and AD Kinghorn. An anthraquinone with potent quinone reductase-inducing activity and other constituents of the fruits of *Morinda citrifolia* (noni). *J. Nat. Prod.* 2005; **68**, 1720-2.
- [4] Jasril, NH Lajis, LY Mooi, MA Abdullah, MA Sukari and AM Ali. Antitumor promoting and antioxidant activities of anthraquinones isolated from the cell suspension culture of *Morinda elliptica*. *AsPac J. Mol. Biol. Biotechnol.* 2003; **11**, 3-7.
- [5] Y Deng, YW Chin, H Chai, WJ Keller and AD Kinghorn. Anthraquinones with quinone reductase-inducing activity and benzophenones from *Morinda citrifolia* (noni) roots. *J. Nat. Prod.* 2007; **70**, 2049-52.
- [6] T Ruksilp, J Sichaem, S Khumkratok, P Siripong and S Tip-pyang. Anthraquinones and an iridoid glycoside from the roots of *Morinda pandurifolia*. *Biochem. Sys. Ecol.* 2011; **39**, 888-92.
- [7] AD Pawlus and AD Kinghorn. Review of the ethnobotany, chemistry, biological activity and safety of the botanical dietary supplement *Morinda citrifolia* (noni). *J. Pharm. Pharmacol.* 2007; **59**, 1587-609.
- [8] O Potterat and M Hamburger. *Morinda citrifolia* (noni) fruit-phytochemistry, pharmacology, safety. *Planta Med.* 2007; **73**, 191-9.
- [9] K Kamiya, W Hamabe, S Harada, R Murakami, S Tokuyama and T Satake. Chemical constituents of *Morinda citrifolia* roots exhibit hypoglycemic effects in streptozotocin-induced diabetic mice. *Biol. Pharm. Bull.* 2008; **31**, 935-8.
- [10] K Kamiya, W Hamabe, S Tokuyama and T Satake. New antraquinone glycosides from the roots of *Morinda citrifolia*. *Fitoterapia* 2009; **80**, 196-9.
- [11] AM Ali, NH Ismail, MM Mackeen, LS Yazan, SM Mohamed, ASH Ho and NH Lajis. Antiviral, cytotoxic and antimicrobial activity of anthraquinones isolated from the root of *Morinda elliptica*. *Pharm. Biol.* 2000; **38**, 298-301.
- [12] Y Zhang, P Talalay, CG Cho and GH Posner. A major inducer of anticarcinogenic protective enzymes from broccoli: Isolation and elucidation of structure. *Proc. Natl. Acad. Sci. USA* 1992; **89**, 2399-403.
- [13] G Rath, M Ndonzao and K Hostettmann. Antifungal anthraquinones from *Morinda lucida*. *Int. J. Pharmacogn.* 1995; **33**, 107-14.
- [14] HL Zhang, QW Zhang, XQ Zhang, WC Ye and YT Wang. Chemical constituents from the root of *Morinda officinalis*. *Chin. J. Nat. Med.* 2010; **8**, 192-5.
- [15] YB Wu, CJ Zheng, LP Qin, LN Sun, T Han, L Jiao, QY Zhang and JZ Wu. Antiosteoporotic activity of anthraquinones from *Morinda officinalis* on osteoblasts and osteoclasts. *Molecules* 2009; **14**, 573-83.
- [16] BS Siddiqui, FA Sattar, F Ahmad and S Begum. Isolation and structural elucidation of chemical constituents from the fruits of

- Morinda citrifolia* Linn. *Arch. Pharm. Res.* 2007; **30**, 919-23.
- [17] GCL Ee, YP Wen, MA Sukari, R Go and HL Lee. A new anthraquinone from *Morinda citrifolia* roots. *Nat. Prod. Res.* 2009; **23**, 1322-9.
- [18] NH Ismail, AM Ali, N Aimi, M Kitajima, H Takayama and NH Lajis. Anthraquinones from *Morinda elliptica*. *Phytochemistry* 1997; **45**, 1723-5.
- [19] K Kamiya, Y Tanaka, H Endang, M Umar and T Satake. New anthraquinine and iridoid from the fruits of *Morinda citrifolia*. *Chem. Pharm. Bull.* 2005; **53**, 1597-9.
- [20] W Xiang, QS Song, H-J Zhang and SP Guo. Antimicrobial anthraquinones from *Morinda angustifolia*. *Fitoterapia* 2008; **79**, 501-4.
- [21] T Akihisa, K Matsumoto, H Tokuda, K Yasukawa, K Seino, K Nakamoto, H Kuninaga, T Suzuki and Y Kimura. Anti-inflammatory and potential cancer chemopreventive constituents of the fruits of *Morinda citrifolia* (noni). *J. Nat. Prod.* 2007; **70**, 754-7.
- [22] BS Siddiqui, FA Sattar, S Begum, T Gulzar and F Ahmad. New anthraquinones from the stem of *Morinda citrifolia* Linn. *Nat. Prod. Res.* 2006; **20**, 1136-44.
- [23] CF Lin, CL Ni, YL Huang, SJ Sheu and CC Chen. Lignans and anthraquinones from the fruits of *Morinda citrifolia*. *Nat. Prod. Res.* 2007; **21**, 1199-204.
- [24] J Takashima, Y Ikeda, K Komiyama, M Hayashi, A Kishida and A Ohsaki. New constituents from the leaves of *Morinda citrifolia*. *Chem. Pharm. Bull.* 2007; **55**, 343-5.
- [25] T Akihisa, K Seino, E Kaneko, K Watanabe, S Tochizawa, M Fukatsu, N Banno, K Metori and Y Kimura. Melanogenesis inhibitory activities of iridoid-hemiterpene-, and fatty acid-glycosides from the fruits of *Morinda citrifolia* (noni). *J. Oleo Sci.* 2010; **59**, 49-57.
- [26] J Schripsema, GP Caprini and D Dagnino. Revision of the structures of citrifolinin A, citrifolinoside, yopaaoside A, yopaaoside B, and morindacin, iridoids from *Morinda citrifolia* L. and *Morinda coreia* Ham. *Org. Lett.* 2006; **8**, 5337-40.
- [27] V Samoylenko, J Zhao, DC Dunbar, IA Khan, JW Rushing and I Muhammad. New constituents from noni (*Morinda citrifolia*) fruit juice. *J. Agric. Food Chem.* 2006; **54**, 6398-402.
- [28] T Kanchanapoom, R Kasai and K Yamasaki. Iridoid and phenolic glycosides from *Morinda coreia*. *Phytochemistry* 2002; **59**, 551-6.
- [29] G Liu, A Bode, WY Ma, S Sang, CT Ho and Z Dong. Two novel glycosides from the fruits of *Morinda citrifolia* (noni) inhibit AP-1 transactivation and cell transformation in the mouse epidermal JB6 cell line. *Cancer. Res.* 2001; **61**, 5749-56.
- [30] P Noiarsa, S Ruchirawat, H Otsuka and T Kanchanapoom. A new iridoid glucoside from the Thai medicinal plant, *Morinda elliptica* Ridl. *J. Nat. Med.* 2006; **60**, 322-4.
- [31] S Sang, M Wang, K He, G Liu, Z Dong, V Badmaev, QY Zheng, G Ghai, RT Rosen and CT Ho. Chemical components in noni fruits and leaves (*Morinda citrifolia* L.). *ACS. Sym. Ser.* 2002; **803**, 134-50.
- [32] XL Yang, MY Jiang, KL Hsieh and JK Liu. Chemical constituents from the seeds of *Morinda citrifolia*. *Chin. J. Nat. Med.* 2009; **7**, 119-22.
- [33] BN Su, AD Pawlus, HA Jung, WJ Keller, JL McLaughlin and AD Kinghorn. Chemical constituents of the fruits of *Morinda citrifolia* (noni) and their antioxidant activity. *J. Nat. Prod.* 2005; **68**, 592-5.
- [34] S Sang, X Cheng, N Zhu, RE Stark, V Badmaev, G Ghai, RT Rosen and CT Ho. Flavonol glycosides and novel iridoid glycoside from the leaves of *Morinda citrifolia*. *J. Agric. Food Chem.* 2001; **49**, 4478-81.
- [35] S Sang, K He, G Liu, N Zhu, M Wang, J Jhoo, Q Zheng, Z Dong, G Ghai, RT Rosen and CT Ho. Citrifolinin A, a new unusual iridoid with inhibition of activator protein-1 (AP-1) from the leaves of noni (*Morinda citrifolia* L.). *Tetrahedron Lett.* 2001; **42**, 1823-5.
- [36] S Sang, G Liu, K He, N Zhu, Z Dong, Q Zheng, RT Rosen and CT Ho. New unusual iridois from the leaves of noni (*Morinda citrifolia* L.) show inhibitory effect on ultraviolet B-induced transcriptional activator protein-1 (AP-1) activity. *Bioorg. Med. Chem.* 2003; **11**, 2499-502.
- [37] K Cimanga, N Hermans, S Apers, SV Miert, H Van den Heuvel, M Claeys, L Pieters and

- A Vlietinck. Complement-inhibiting iridoids from *Morinda morindoides*. *J. Nat. Prod.* 2003; **66**, 97-102.
- [38] S Tamura, BK Kubata, Syamsurizal, S Itagaki, T Horii, MK Taba and N Murakami. New anti-malarial phenylpropanoid conjugated iridoids from *Morinda morindoides*. *Bioorg. Med. Chem. Lett.* 2010; **20**, 1520-3.
- [39] S Sang, X Cheng, N Zhu, M Wang, JW Jhoo, RE Stark, V Badmaev, G Ghai, RT Rosen and CT Ho. Iridoid glycosides from the leaves of *Morinda citrifolia*. *J. Nat. Prod.* 2001; **64**, 799-800.
- [40] S Sang, K He, G Liu, N Zhu, X Cheng, M Wang, Q Zheng, Z Dong, G Ghai, RT Rosen and CT Ho. A new unusual iridoid with inhibition of activator protein-1 (AP-1) from the leaves of *Morinda citrifolia* L. *Org. Lett.* 2001; **3**, 1307-9.
- [41] YJ Lim, EH Lee, TH Kang, SK Ha, MS Oh, SM Kim, TJ Yoon, C Kang, JH Park and SY Kim. Inhibitory effects of arbutin on melanin biosynthesis of  $\alpha$ -melanocyte stimulating hormone-induced hyperpigmentation in cultured brownish guinea pig skin tissues. *Arch. Pharm. Res.* 2009; **32**, 367-73.